



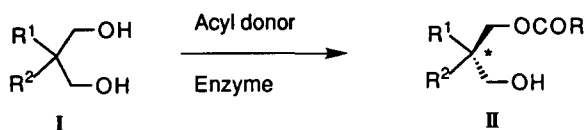
## Enzyme-catalyzed Asymmetrization of 2,2-Disubstituted 1,3-Propanediols Using 1-Ethoxyvinyl Esters

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**Abstract:** The lipase-catalyzed asymmetrization of the prochiral 2,2-disubstituted 1,3-propanediols **1**, **6**, and **9** was studied using various types of 1-ethoxyvinyl esters **2**. Among them, the use of the benzoate **2f** provided good chemical and optical yields of the products **3**, **7**, and **10** bearing chiral quaternary carbon centers which are fairly stable against racemization. © 1997 Elsevier Science Ltd.

The synthetic importance as well as chemical interest for the construction of optically active quaternary carbon centers have widely been recognized.<sup>1</sup> Among the increasing number of novel methods for this topic, the enzymatic differentiation of the prochiral hydroxyl groups of the 2,2-disubstituted 1,3-propanediols **I** leading to the chiral compounds **II** must be attractive (Scheme 1). However, this approach has rarely been reported,<sup>2-5</sup> although many successful examples are known for the similar asymmetrization of 2-monosubstituted 1,3-propanediols (**I** where  $R^2 = H$ ).<sup>6</sup> This seems to be due to the low reactivity of the sterically congested substrates **I** and the easy racemization of the products **II** through acyl group migration.



Scheme 1

Very recently, we have disclosed a novel enzymatic resolution of the alcohols using the 1-ethoxyvinyl ester (EVE) as an acyl donor.<sup>7</sup> In contrast to the well known method using vinyl esters, our method features the formation of ethyl acetate as a single side-product which does not disturb the enzymatic reactions, therefore, it allows the reliable use of a variety of enzymes.<sup>8</sup> It also shows comparative to or higher reactivity than the usual method and its asymmetric efficiency is similarly high. We now demonstrate additional advantages of our method in the above mentioned enzymatic asymmetrization of prochiral 2,2-disubstituted 1,3-propanediols **I**, in which ready availability of EVEs having various kinds of acyl groups and their high reactivity led us to overcome these problems.

At first, we attempted the reaction of the diol **1** with ethoxyvinyl acetate **2a** (3 equiv.) in the presence of several lipases (Amano PS, AK, AY, PPL, Toyobo LIP) in hexane or *i*-Pr<sub>2</sub>O at 30 °C. However, the maximum ee of the desired monoacetate **3a** was 27% which was obtained using Amano AY (from *Candida rugosa*) in *i*-Pr<sub>2</sub>O (Run 1 in Table 1).

We then investigated other acyl group donors **2b-i** instead of **2a**. The reagents **2b-i** were prepared in good yields by the addition of the corresponding carboxylic acid to ethoxyacetylene catalyzed by [RuCl<sub>2</sub>(*p*-

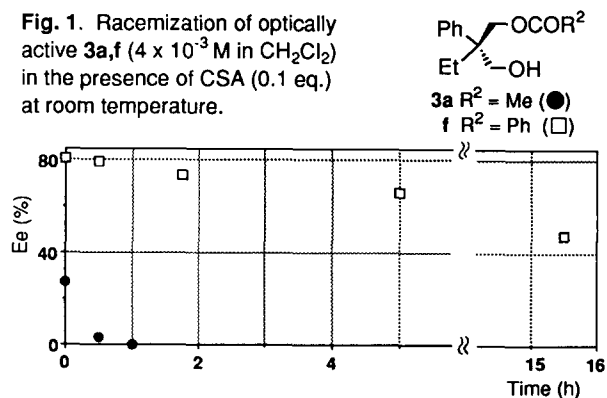
cymene)]<sub>2</sub>.<sup>9</sup> Similar reactions of **1** with **2b-i** were carried out in the presence of Amano AY in *i*-Pr<sub>2</sub>O, and the results are summarized in Table 1.

**Table 1.** Lipase-catalyzed asymmetrization of the 1,3-diols **1** and **6** using various types of **2**

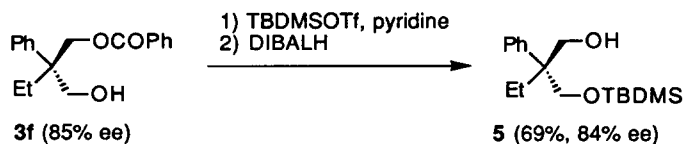
Run	Diol	2, R <sup>2</sup> =	Reaction time	Monoester <b>3,7</b>		Diester <b>4,8</b>
				Isolated yield, % <sup>a</sup>	Ee, % <sup>b</sup>	Isolated yield, % <sup>a</sup>
1	<b>1</b>	<b>a</b> Me	4 d	<b>3a</b> 66	27	<b>4a</b> --
2	<b>1</b>	<b>b</b> CH <sub>2</sub> Cl	7 d	<b>b</b> 36	1 <sup>c</sup>	<b>b</b> 64
3	<b>1</b>	<b>c</b> C <sub>3</sub> H <sub>7</sub>	5 h	<b>c</b> 86	63	<b>c</b> 14
4	<b>1</b>	<b>d</b> C <sub>7</sub> H <sub>15</sub>	4 d	<b>d</b> 85	36	<b>d</b> 12
5	<b>1</b>	<b>e</b> C <sub>11</sub> H <sub>23</sub>	21 d	<b>e</b> 78	0 <sup>c</sup>	<b>e</b> --
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6	<b>1</b>	<b>f</b> C <sub>6</sub> H <sub>5</sub>	6 h	<b>f</b> 90	81	<b>f</b> --
7	<b>1</b>	<b>f</b> C <sub>6</sub> H <sub>5</sub>	18 h	<b>f</b> 39	91	<b>f</b> 61
8	<b>1</b>	<b>g</b> 4-MeO-C <sub>6</sub> H <sub>4</sub>	8.5 d	<b>g</b> 50	69	<b>g</b> 33
9	<b>1</b>	<b>h</b> 4-NO <sub>2</sub> -C <sub>6</sub> H <sub>4</sub>	13 d	<b>h</b> 64	53	<b>h</b> d
10	<b>1</b>	<b>i</b> 2,6-diMe-C <sub>6</sub> H <sub>3</sub>	30 d	<b>i</b> <5	--	<b>i</b> --
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11	<b>6</b>	<b>a</b> Me	8 d	<b>7a</b> 46	1	<b>8a</b> ca.10
12	<b>6</b>	<b>f</b> C <sub>6</sub> H <sub>5</sub>	3 d	<b>f</b> 71	84	<b>f</b> 26

a, Purified by SiO<sub>2</sub>-column chromatography. b, Determined by Daisel CHIRALCEL OD (hexane-*i*-PrOH). c, Determined after transformation into **5** by a method similar to that shown in Scheme 2. d, Not determined.

Several aspects are noteworthy. First, the benzoate **2f** was the most effective in terms of high chemical and optical yields as well as high reactivity (Run 6).<sup>10</sup> A prolonged reaction gave a slightly better ee of **3f**, although its yield was decreased (Run 7). Second, the other aryl reagent, *p*-methoxybenzoate **2g** (Run 8), was in the second best class along with the aliphatic butyryl reagent **2c** (Run 3). These effective results of the aryl donors **2f,g** are outstanding because the successful use of benzoyl donors for the enzymatic reactions have been reported in limited cases.<sup>11</sup> Moreover, the product **3f** having a benzoyl group was found to be relatively stable against the racemization compared to the acetate **3a**. A comparison of the stability of these two products under acidic conditions [0.1 equiv. of camphor sulfonic acid (CSA), CH<sub>2</sub>Cl<sub>2</sub>, room temperature] is shown in Fig. 1.<sup>12</sup>



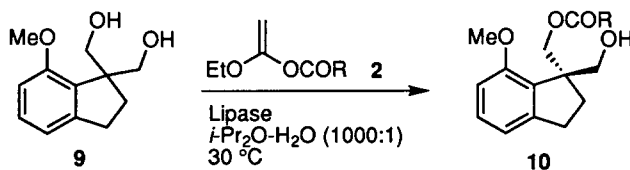
A preliminary transformation of the functional group of **3f** into the silyl ether **5** was attained without loss of its optical purity (Scheme 2).



Similarly, asymmetric reduction of **6** using **2f** and Amano AY gave the chiral product **7f** in 71% yield with 84% ee,<sup>10</sup> while a similar reaction using **2a** was not satisfactory (Table 1; Runs 11 and 12).

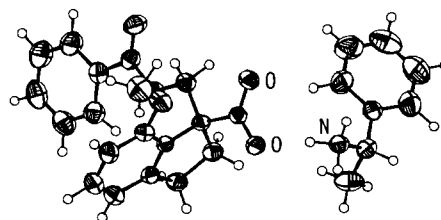
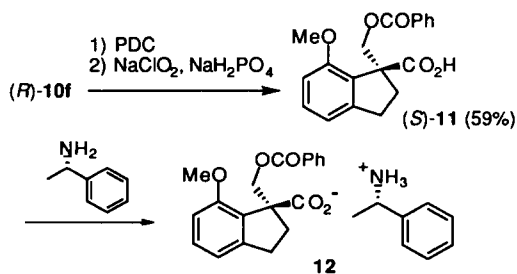
This method was also applied to the more hindered diol **9** (Table 2). The use of the benzoate **2f** gave 72% ee **10f** in 50% isolated yield (Run 3), while the use of **2a** or **2c** was again insufficient (Runs 1,2). A better result (82% yield, 71% ee) was obtained using a similar lipase MY (Meito Sangyo) obtained from *Candida rugosa* (Run 4). The product **10f** [71% ee,  $[\alpha]_D^{24}$  -39.9 (c 0.4, CHCl<sub>3</sub>)] was converted to the carboxylic acid **11** (59% yield). An enantiomerically pure **11** was obtained through formation of a crystalline salt **12** with (*S*)-1-phenethylamine followed by recrystallization from EtOH (Scheme 3). In contrast to the cases of **3f** and **7f**, the absolute stereochemistry of the chiral center of **10f** was disclosed to be *R*, because the X-ray crystallographic analysis of **12** unambiguously showed that the chiral center of **11** was *S*.

**Table 2.** Lipase-catalyzed asymmetric reduction of the 1,3-diol **9**.



Run	Lipase	2, R =	Reaction time	10	
				Isolated yield, % <sup>a</sup>	Ee, % <sup>b</sup>
1	Amano AY	a Me	25 d	a 41	10
2	Amano AY	c C <sub>3</sub> H <sub>7</sub>	25 d	c 37	10
3	Amano AY	f C <sub>6</sub> H <sub>5</sub>	4 d	f 50	72
4	Lipase MY <sup>c</sup>	f C <sub>6</sub> H <sub>5</sub>	4 d	f 82	71
5	Lipase OF <sup>c</sup>	f C <sub>6</sub> H <sub>5</sub>	4 d	f 67	58

a, Purified by SiO<sub>2</sub>-column chromatography. b, Determined by Daicel CHIRALCEL OD (hexane-*i*-PrOH). c, From Meito Sangyo.



The X-Ray crystallographic structure of the salt **12**.

